

Comparative evaluation of the antimicrobial efficacy of aloe vera tooth gel and two popular commercial toothpastes: An *in vitro* study

Dilip George, MDS - Sham S. Bhat, MDS - Beena Antony, PhD

Aloe vera (Aloe barbadensis Miller) has been suggested for a wide variety of ailments but its use in dentistry is limited. This article reviews the uses of the plant and describes an *in vitro* investigation that compared the antimicrobial effectiveness of aloe vera tooth gel with two popular, commercially available dentifrices. The preliminary results showed that aloe vera tooth gel and the toothpastes were equally effective against Candida albicans,

Streptococcus mutans, Lactobacillus acidophilus, Enterococcus faecalis, Prevotella intermedia, and Peptostreptococcus anaerobius. Aloe vera tooth gel demonstrated enhanced antibacterial effect against *S. mitis*.

Received: November 29, 2007 Accepted: February 8, 2008

he success of any toothpaste, in part, lies on its ability to eliminate pathogenic oral microflora. Fluoride dentifrices have been widely used all over the world and extensive research has established their abilities in terms of caries resistance.¹

The gel or mucilage from *Aloe barbadensis Miller* (otherwise known as *aloe vera*) is a convenient homegrown remedy that can be used both as a moisturizing agent and for treating minor burns and skin abrasions. Aloe vera is a cactuslike plant that actually is part of the lily family. There are more than 300 varieties of the aloe plant but the *Aloe barbadensis* variety exhibits the best medicinal properties.

Modern use of aloe vera was first documented in the 1930s to heal radiation burns.² Aloe vera juice taken internally has been shown to have various beneficial effects on the body.³⁻⁶

The efficacy of *Aloe barbadensis Miller* increases when the plant is harvested after three years of

growth but its nutritive potency decreases after 12 years of growth.7 Aloe gel will lose its complete potency if it is exposed to sunlight for more than two hours, as it is easily oxidized; consequently, it is necessary to stabilize it under pharmaceutical standards for ready use and longer shelf life. Non-profit organizations like the International Aloe Science Council have set standards for aloe vera approval and give their seal of quality for aloe products. Such products are more beneficial, since the seal is given to only those products with established therapeutic benefits.7

This *in vitro* evaluation compared the antimicrobial activity of an aloe vera tooth gel and two commercially popular, locally available toothpastes. These toothpastes were tested against seven pathogenic microorganisms that frequently dominate the oral microbiota. The results are intended to show the relative antimicrobial effectiveness of each dentifrice against each particular species.

Materials and methods

To demonstrate antimicrobial activity, this study utilized an aloe vera tooth gel (Forever Bright, Forever Living Products, Scottsdale, AZ; 888.440.2563), known as Toothpaste A, and two commercial, locally available toothpastes: Pepsodent (Unilever, Englewood Cliffs, NJ; 201.894.7660) and Colgate (Colgate-Palmolive, Canton, MA; 800.821.2880), known as Toothpastes B and C, respectively.

This study used freeze-dried stock culture of the reference strains of Streptococcus mutans, Candida albicans, Lactobacillus acidophilus, S. mitis, Enterococcus faecalis, Prevotella intermedia, and Peptostreptococcus anaerobius. The organisms were cultured in trypticase soy broth and transferred to the selective media to revive from the stock.

The organisms employed in the study were cultured in their respective selective media. For example, *S. mutans* was cultured in Mitis Salivarius Bacitracin Agar (Gold's Media), *C. albicans* in Sabouraud's Dextrose

Agar, *L. acidophilus* in Rogosa SL Agar, *S. mitis* in Mitis-Salivaris Agar, *E. faecalis* in Mac Conkey's Agar, and both *Prevotella intermedia* and *Peptostreptococcus anaerobius* in Neomycin Blood Agar.⁸

The purity of each test strain was checked during each trial by using subculture, Gram's stain, and colony morphology. Antimicrobial susceptibility was checked by using the ditch method. The Muller Hinton Agar was used to demonstrate the antibacterial effect on aerobes, while Wilkins Chalgren Blood Agar was used for anaerobes and Sabouraud's Dextrose Agar for *Candida*.

Three wells (4 mm in diameter and 3 mm deep) were made using a sterile metallic template, with a rubber teat in each plate. The inoculums were prepared and adjusted to 0.5 McFarland turbidity standards, according to National Committee on Clinical Laboratory Standards (NCCLS) guidelines.10 The agar plates were streaked with the respective stock culture microorganisms. Using a sterile spoon excavator, the toothpastes were dispersed into the wells. At that point, the plates were incubated at 37°C for 48 hours in the respective environments—that is, E. faecalis and C. albicans in the incubator, S. mutans and S. mitis in the candle jar, and *L. acidophilus*, Prevotella intermedia, and Peptostreptococcus anaerobius in an anaerobic jar (Hi Anaerobic System-Mark II with Anaerobic Hi Gas Pack, Hi Media Laboratories, Mumbai, India; 91.022.2500.0970), which works on the principle of gas generated from chemicals.

After incubation, zones of inhibition (that is, locations where no growth of bacteria was present) were examined around the wells that contained the dentifrice. These appeared as a clear, circular halo surrounding the wells. Diameters

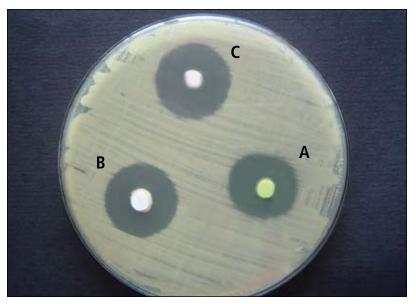


Fig. 1. Zone of inhibition as observed in *Candida albicans* for toothpastes A, B, and C, using Sabouraud's Dextrose Agar.

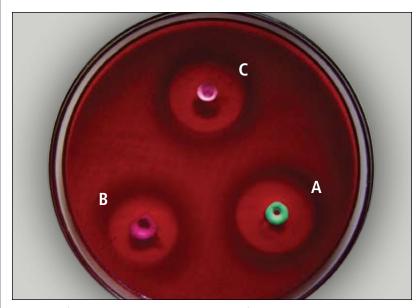


Fig. 2. Zone of inhibition observed in *Prevotella intermedia* for toothpastes A, B, and C, using Wilkins Chalgren Blood Agar.

of the zones were measured with a Hi Antibiotic Zone Scale (Hi Media Laboratories). The mean diameter of the well's measurements (in mm) represented the inhibition value of the tested product. No attempt was made to obscure the identity of the test agents. The test was repeated six times in triplicate to overcome any

Table. Mean diameter of the zone of inhibition obtained after 48 hours of incubation.

		N	Mean	SD	Н	р
	S. mutans				4.18	0.124 (not significant)
	Α	6	15.8333	0.75277		
	В	6	15.5000	1.04881		
	C	6	16.8333	1.16905		
	C. albicans				5.48	0.058 (not significant)
	Α	6	24.0000	0.823666		
	В	6	25.0000	0.89443		
	C	6	23.6667	1.03280		
	L. acidophilus				0.87	0.647 (not significant)
	Α	6	23.1667	3.54495		
	В	6	23.8333	3.37145		
	C	6	22.8333	3.06050		
	S. mitis				6.76	0.034 (significant)
	Α	6	17.0000	3.16228		
	В	6	14.6667	1.50555		
	C	6	14.3333	1.75119		
	E. faecalis				0.84	0.659 (not significant)
	Α	6	22.3333	2.06559		
	В	6	23.0000	2.19089		
	C	6	23.3333	2.06559		
	Prevotella intermedia				4.83	0.09 (not significant)
	Α	6	21.3333	1.03280		
	В	6	22.1667	0.98319		
	C	6	20.8333	0.75277		
Peptostreptococcus anaerobius				0.07	0.968 (not significant)	
	Α	6	21.6667	0.81650		
	В	6	21.5000	1.37840		
	C	6	21.6667	1.36626		

technical errors that might have occurred during a single attempt. The Kruskall Wallis Test was utilized, with SPSS Version 14 software used to analyze the results.

Results

Results of this preliminary in vitro study demonstrated that aloe vera tooth gel was equally effective as Toothpastes B and C for controlling all of the organisms in the study. All three toothpastes showed

maximum antimicrobial activity against C. albicans and all anaerobes in vitro.

Compared to the Toothpastes B and C, Toothpaste A demonstrated an increased antibacterial effect against *S. mitis* (p = 0.034). The table lists the zone of inhibition obtained from each toothpaste after 48 hours.

Discussion

A review of the literature suggested that the potential of using aloe

vera for oral hygiene had not been evaluated prior to this study. The antibacterial, antifungal, and antiviral properties of aloe vera have been established; in addition, it reduces inflammation and pain and aids in healing.

The antimicrobial effects of aloe vera have been attributed to the plant's natural anthraquinones: aloe emodin, aloetic acid, aloin, anthracine, anthranol, barbaloin, chrysophanic acid, ethereal oil, ester of cinnamonic acid, isobarbaloin, and resistannol.11 In relatively small concentrations together with the gel fraction, these anthraquinones provide analgesic, antibacterial, antifungal, and antiviral activity; in high concentrations, they can be toxic.¹² Saponins, which contain glycosides, are soapy substances that have both cleansing and antiseptic properties. 13-15

Acemannan, a complex mannose carbohydrate derived from the aloe vera plant, has an inherent stickiness/viscosity, which makes it ideal for denture adhesive formulations. A 1998 study reported that acemannan formulations of 150:1 (containing 0.05% benzalkonium chloride, 0.1% methylparaben, and 0.01% hyamine 1622) exhibited ideal adhesive strength and pH and minimal cytotoxicity.¹⁶

The organisms employed in the present study include both the normal flora and pathogens of the oral cavity. S. mutans has been strongly associated with the initiation of caries, while there is a correlation between Lactobacilli and the further development of carious lesions.¹⁷ A 1995 study by Bai et al demonstrated a high Candida count in children with insulin-dependent diabetes mellitus, a condition associated with symptoms like dry mouth, burning sensations, and painful fissures.¹⁸ E. faecalis has been associated with

re-infection and subsequent failure of endodontically treated teeth.¹⁹

Anaerobes are a significant part of orodental flora. Their role in periodontal disease and root canal infection is well-established, as is their role as foci for disseminated infectious disease. *Prevotella intermedia* were predominant among the anaerobes recovered from these periodontal infections, which meant these particular organisms were appropriate for the present study.²⁰

The majority of the antimicrobial effects of commercially available toothpastes can be attributed to their fluoride content, in the form of sodium monoflourophosphate (a concentration of 500–1,000 ppm). The aloe vera tooth gel used in the present study has no added fluoride content but still exerts almost an equal amount of antimicrobial activity.

Conclusion

This preliminary *in vitro* study demonstrated that aloe vera tooth gel was as effective as two commercially popular toothpastes in controlling all of the organisms used in the study. In addition, the gel demonstrated superior antibacterial effect against *S. mitis* despite the absence of additional fluoride. However, to guarantee these results and the effectiveness of these tooth

care products, additional long-term clinical trials should be performed that incorporate more isolates from clinical samples.

Disclaimer

The authors have no relationship with any of the manufacturers cited in this article.

Author information

Dr. George is a senior lecturer/ assistant professor, Department of Pedodontics and Preventive Dentistry, Pushpagiri College of Dental Sciences, Tiruvallam, Kerala, India. Dr. Bhat is a professor and head, Department of Pedodontics and Preventive Dentistry, Yenepoya Dental College Hospital, Mangalore, Karnataka, India. Dr. Antony is a professor, Department of Microbiology, Father Muller Medical College Hospital, Mangalore, Karnataka, India.

References

- Itthagarun A, Wei SH. Analysis of fluoride ion concentrations and *in vitro* fluoride uptake from different commercial dentifrices. Int Dent J 1996;46(4);357-361.
- Collins CE. Alvagel as a therapeutic agent in the treatment of roentgen and radium burns. Radiol Rev Chicago Med Rec 1935;57:137-138.
- 3. PDR for herbal medicines. Montvale, NJ: Medical Economics Company;1998:631.
- 4. Red book, 2004. Montvale, NJ: Thomson Healthcare; 2004:53.
- 5. Tyler V. The honest herbal: A sensible guide to the use of herbs and related remedies, ed. 3.

- New York: Pharmaceutical Products Press;1993: 25-28
- Krinsky DL, Hawkins EB, Pelton R, Willis NA, Lavalle JB. Natural therapeutics pocket guide, ed. 2. Cleveland: Lexi-Comp, Inc.;2003:379.
- 7. Venkatrama EV. The miracle worker. New Indian Express 2005;November 22:3.
- Gold DG, Jordan HV, Van Houte J. A selective medium for *Streptococcus mutans*. Arch Oral Biol 1973;18:1357-1364.
- Ananthanarayanan R, Panicker CKJ. Text book of microbiology, ed. 7. Hyderabad, India: Orient Black Swan; 2005:628.
- National Committee for Clinical Laboratory Standards. Performance standards for antimicrobial disc susceptibility tests; approved standard, ed. 8. Wayne, PA: National Committee for Clinical Laboratory Standards; 2003:9.
- 11. Wynn RL. Aloe vera gel: Update for dentistry. Gen Dent 2005;53(1):6-9.
- 12. Davis RH. Aloe vera: A scientific approach. New York: Vantage Press; 1997.
- 13. Plaskett LG. The health and medical use of aloe vera. Tacoma, WA: Life Sciences Press; 1996.
- 14. Coats BC. The silent healer: A modern study of aloe vera. Garland, TX: B.C. Coats; 1979.
- Gage D. Aloe vera: Nature's soothing healer. Rochester, VT: Healing Arts Press; 1996.
- Tello CG, Ford P, Iacopino AM. In vitro evaluation of complex carbohydrate denture adhesive formulations. Quintessence Int 1998;29(9):585-593.
- Zickert I, Emilson CG, Krasse B. Streptococcus mutans, lactobacilli and dental health in 13-14year-old Swedish children. Community Dent Oral Epidemiol 1982;10(2):77-81.
- Bai KY, Reddy CD, Abu-Talib SH. Oral candidial carriage in young insulin dependent diabetics. J Ind Pedo Prev Dent 1995;13(1):20-23.
- Kayaoghu G, Orstavik D. Virulence factors of *Enterococcus faecalis*: Relationship to endodontic disease. Crit Rev Oral Biol Med 2004;15(5): 308-320.
- Newman MG. Anaerobic oral and dental infections. Rev Infect Dis 1984;6 Suppl 1:S107-S114.

Published with permission by the Academy of General Dentistry. © Copyright 2009 by the Academy of General Dentistry. All rights reserved.